

# Hierarchically Restructured Antibacterial Electrodes for Neural Interfaces: Electrochemical and Microstructural Evolution under Extended Cycling

Kriti Panchal, Wesley Seche, Henna Khosla, Gang Feng, Jacob Elmer, Gregory A. Caputo, Steven J. May, Ekaterina Pomerantseva,\* and Shahram Amini\*



Cite This: <https://doi.org/10.1021/acsami.5c21727>



Read Online

ACCESS |



Metrics & More



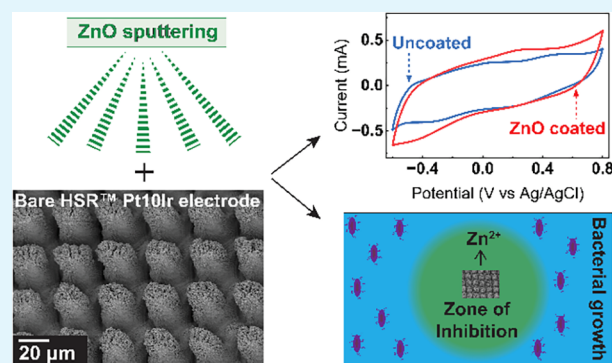
Article Recommendations



Supporting Information

**ABSTRACT:** Hierarchically restructured platinum–iridium electrodes offer high electrochemical performance for neurostimulation and cardiac rhythm management devices but require added antibacterial functionality to reduce postsurgical infection risks. In this work, electrochemically active antibacterial platinum–iridium electrodes were developed using a two-step process. First, the electrodes were restructured using a femtosecond laser hierarchical surface restructuring. In the second step, reactive magnetron sputtering from a pure zinc target in an Ar/O<sub>2</sub> gas mixture was employed to deposit antibacterial zinc oxide (ZnO) thin films onto the hierarchical surface structure of the electrodes, thereby imparting antibacterial properties. X-ray diffraction and X-ray photoelectron spectroscopy confirmed the formation of ZnO. The electrochemical performance of the electrodes increased with the ZnO film deposition time. This enhancement is attributed to the nonconformal nature of the ZnO layer over the complex electrode topography, as revealed by scanning electron microscopy (SEM). SEM imaging combined with energy-dispersive spectroscopy (EDS) mapping after electrochemical cycling revealed the gradual dissolution of ZnO into the electrolyte and the recrystallization of ZnO on the electrode surface after 1,500 cyclic voltammetry (CV) cycles (24 h), likely due to the confined electrolyte environment. Electrodes coated with ZnO films exhibited significant antibacterial activity against *Escherichia coli* and *Staphylococcus aureus* bacterial strains *in vitro*. The findings of this work highlight a promising strategy for developing multifunctional, electrochemically active antibacterial electrodes for next-generation neural interfacing electrodes.

**KEYWORDS:** hierarchical surface restructuring, antibacterial coatings, neural interfacing, electrodes, platinum–iridium, zinc oxide films, electrochemically active surface area



## 1. INTRODUCTION

Long-term implantable medical devices represent a rapidly advancing frontier in modern medicine, with their development requiring precise optimization of material properties and functional parameters to ensure long-term efficacy and reliability. Among these technologies, neurostimulation<sup>1–4</sup> and cardiac rhythm management (CRM)<sup>5,6</sup> devices play a critical role in the treatment of a wide range of neurological and cardiac disorders by delivering targeted electrical stimulation to specific sites within the central or peripheral nervous system or the myocardium, thereby modulating biological activity through inhibition, excitation, or alteration of native signal pathways.

This stimulation is achieved by transferring externally generated electrical signals from a neurostimulator or an implantable pulse generator (IPG) via a lead to an implantable electrode or microelectrode array, thereby inducing controlled changes in neural activity.<sup>7</sup> Among these system components,

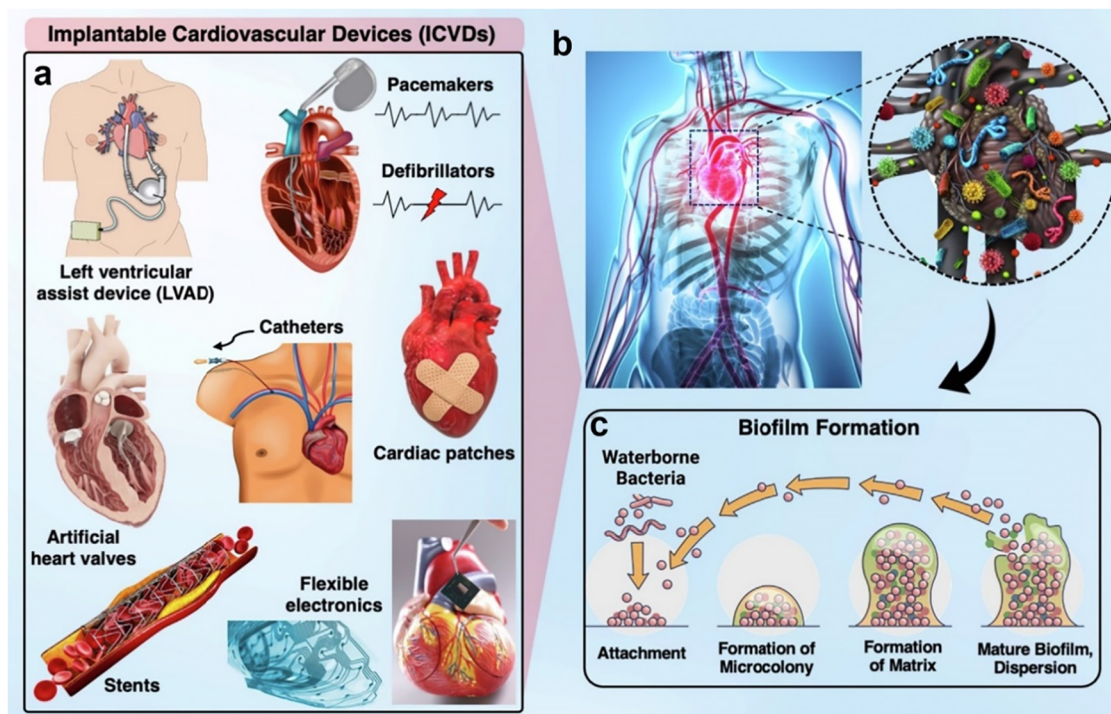
electrodes and microelectrode arrays serve as the primary interface between the device and the biological environment, playing a pivotal role in ensuring the precise, efficient, and safe delivery of electrical stimuli. A critical requirement for electrodes used in neural interfaces such as neurostimulation and CRM applications is their electrochemical performance, which governs both the fidelity of neural modulation (or cardiac pacing) and the long-term stability of the device.

Hierarchically restructured platinum–iridium (Pt10Ir) electrodes, fabricated using femtosecond-laser hierarchical surface restructuring (HSR) technology, have recently demonstrated

**Received:** October 29, 2025

**Revised:** February 26, 2026

**Accepted:** March 2, 2026



**Figure 1.** (a) and (b) Examples of implantable cardiovascular devices prone to biofilm-related infections, including stents, pacemakers, defibrillators, heart valves, cardiac patches, and flexible electronics. (c) Schematic of biofilm formation mechanism. Reprinted from Current Opinion in Biomedical Engineering,<sup>17</sup> Vol. 23, E. Mostafavi, A.K. Dubey, B. Walkowiak, A. Kaushik, S. Ramakrishna, L. Teodori, “Antimicrobial surfaces for implantable cardiovascular devices,” p. 100406, © 2022 Elsevier, with permission.

56 exceptional electrochemical performance in previous studies,<sup>8–10</sup> making them highly promising for advanced neural  
57 interfacing applications. Their enhanced electrochemical  
58 activity is attributed to the engineered hierarchical surface  
59 topography, consisting of primary micropillar structures  
60 approximately 10–20  $\mu\text{m}$  in height, overlaid with secondary  
61 nanoscale features ranging from a few nanometers to several  
62 hundred nanometers. This multiscale architecture significantly  
63 increases the electrochemical surface area (ESA) while  
64 preserving geometric compactness, thereby improving both  
65 the charge transfer efficiency and signal fidelity. This  
66 hierarchical architecture significantly enhances the ESA  
67 without increasing the overall electrode footprint, resulting in  
68 marked improvements in both specific capacitance and charge  
69 storage capacity.<sup>8,10</sup> However, despite their superior electro-  
70 chemical characteristics, HSR-processed electrodes inherently  
71 lack antibacterial properties, necessitating additional surface  
72 modifications to impart antimicrobial functionality without  
73 compromising their electrochemical performance.

74  
75 Antibacterial functionality remains a critical design consid-  
76 eration across all implantable medical devices. Following  
77 implantation, these devices are often recognized as foreign  
78 bodies by the immune system, triggering inflammatory  
79 responses that can facilitate bacterial and biofilm coloniza-  
80 tion.<sup>11</sup> Despite the use of immunosuppressive strategies,  
81 postimplantation infections occur in approximately 20% of  
82 patients,<sup>12–16</sup> often resulting in serious complications and  
83 postsurgical hospitalizations. Figure 1 illustrates representative  
84 examples of implantable devices commonly used in clinical  
85 practice, along with a schematic illustration of the biofilm  
86 formation mechanism that can occur following implantation.

## 2. OBJECTIVES

This study focuses on addressing the critical need to balance 87  
88 electrochemical performance and antibacterial activity in  
89 implantable electrodes for neural interfacing applications.  
90 One promising strategy for achieving this balance is the  
91 incorporation of antibacterial surface coatings.<sup>18,19</sup> Among the  
92 materials investigated, zinc (Zn), silver (Ag), and copper (Cu)-  
93 based coatings have been extensively studied for their broad-  
94 spectrum antibacterial efficacy against various bacterial  
95 strains.<sup>17,20–22</sup> Of these, zinc oxide (ZnO) has emerged as a  
96 particularly attractive candidate for implant applications, owing  
97 to its multiple antibacterial mechanisms and favorable  
98 compatibility with biomedical environments.<sup>23–28</sup>

99 There are two possible mechanisms suggested for the  
100 antibacterial activity of ZnO.<sup>20,28</sup> The first mechanism is the  
101 generation of oxygen radicals and hydrogen peroxide ( $\text{H}_2\text{O}_2$ )  
102 from the defect sites on the ZnO surface.<sup>29–31</sup> When defect-  
103 rich ZnO is activated by UV or visible light, electron–hole  
104 pairs ( $e^-/h^+$ ) form. The holes split  $\text{H}_2\text{O}$  present in the  
105 surrounding biological media into  $\text{OH}^-$  and  $\text{H}^+$ , while  $\text{O}_2$  is  
106 reduced to superoxide radicals ( $\text{O}_2^-$ ), which further react to  
107 generate  $\text{H}_2\text{O}_2$ .<sup>29</sup> The produced  $\text{H}_2\text{O}_2$  can penetrate bacterial  
108 cell membranes and induce cell death.<sup>25,29,32</sup> The second  
109 mechanism is causing damage to the bacterial cell wall due to  
110 electrostatic interactions of the dissolved  $\text{Zn}^{2+}$  ions from the  
111 ZnO surfaces.<sup>23,24,27,28,33</sup>

112 Specifically, previous studies have shown that ZnO is  
113 selectively toxic to both Gram-negative and Gram-positive  
114 bacteria than healthy human cells,<sup>34,35</sup> making it a leading  
115 choice for an antibacterial coating. Although the antibacterial  
116 efficacy of ZnO is relatively lower compared to silver (Ag) and  
117 copper (Cu), the cytotoxicity of  $\text{Zn}^{2+}$  ions is significantly  
118 reduced across a wider concentration range,<sup>22,23,26</sup> making

119 ZnO a safer and more biocompatible alternative for  
120 applications in long-term implantable biomedical devices.

121 Despite their antibacterial potential, ZnO coatings must also  
122 demonstrate compatibility with the electrochemical perform-  
123 ance requirements of HSR-Pt10Ir electrodes, which are used in  
124 neural interfacing applications. While ZnO is moderately  
125 conductive and offers advantages such as cost-effective  
126 fabrication and broad-spectrum antibacterial efficacy, its  
127 electrochemical behavior—particularly when applied to a  
128 hierarchically restructured electrode surface—remains largely  
129 unexplored. Therefore, this study investigates the electro-  
130 chemical and microstructural evolution of antibacterial ZnO  
131 coatings deposited on HSR-processed Pt10Ir electrodes with  
132 the aim of evaluating their viability for dual-function neural  
133 interfacing and CRM applications.

134 In this study, ZnO coatings were deposited via reactive DC  
135 magnetron sputtering onto the HSR-Pt10Ir electrodes. Two  
136 deposition durations of 5 and 60 min were selected to evaluate  
137 the effect of coating thickness on surface and electrochemical  
138 properties of ZnO-coated HSR-Pt10Ir electrodes. Compre-  
139 hensive physical characterization of the ZnO-coated electrodes  
140 was conducted to assess their structural, compositional, and  
141 morphological features.

142 Electrochemical measurements were performed to evaluate  
143 the impact of ZnO deposition on the performance of HSR-  
144 Pt10Ir electrodes in a physiological saline solution, simulating  
145 the ionic environment of bodily fluids. Antibacterial activity  
146 was measured against Gram-positive (*S. aureus*) and Gram-  
147 negative (*E. coli*) bacteria. We show that ZnO coatings applied  
148 via reactive DC magnetron sputtering to HSR-Pt10Ir electro-  
149 des can impart antibacterial functionality to implantable Pt10Ir  
150 electrodes while preserving the electrochemical activity  
151 essential for long-term clinical performance.

### 3. MATERIALS AND METHODS

#### 3.1. Electrode Fabrication

152 The electrode fabrication process consisted of two main steps: (i)  
153 hierarchical surface restructuring (HSR) of Pt10Ir electrodes and (ii)  
154 ZnO layer deposition. The full experimental details of the hierarchical  
155 surface restructuring (HSR) process have been discussed in previous  
156 reports.<sup>8–10,36</sup> Briefly, it was performed using a femtosecond laser  
157 system (Coherent StarFemto, 1030 nm, 300 fs pulses) under ambient  
158 conditions. Flat (unrestructured) Pt10Ir foils (0.3 mm thick) were  
159 used as the electrode material and laser-cut into 6 mm diameter discs  
160 for electrochemical measurements, 10 mm diameter discs for  
161 antibacterial testing, and 10 × 10 mm squares for physical  
162 characterization. Surface patterns were generated by using the Visual  
163 Laser Marker software integrated with motion and beam control. This  
164 process enabled the formation of reproducible micro/nanostructured  
165 features, enhancing the surface area and electrode functionality.

166 A planar DC magnetron sputtering system with a zinc target was  
167 employed for reactive sputtering deposition in an oxidizing  
168 atmosphere. The deposition utilized an oxygen flow rate of 40  
169 sccm, an argon flow rate of 50 sccm, a sputtering pressure of 4 mTorr,  
170 a target-to-substrate spacing of 4 mm, and an RF power of 250 W. All  
171 depositions were performed at ambient substrate temperature. The  
172 substrates used included planar silicon wafers and HSR-Pt10Ir  
173 electrodes. To enhance ZnO film adhesion, a primary zinc layer  
174 was deposited in a pure argon atmosphere for 1 min before  
175 introducing oxygen for ZnO deposition. ZnO films were deposited  
176 for 5 and 60 min to examine representative thin and thick coating  
177 regimes and their influence on the electrochemical behavior of HSR-  
178 Pt10Ir electrodes. The resulting ZnO-coated HSR-Pt10Ir electrodes  
179 are referred to as HSR-Pt10Ir-ZnO-5 and HSR-Pt10Ir-ZnO-60,

corresponding to deposition durations of 5 and 60 min, respectively, 180  
throughout the paper. 181

#### 3.2. Physical Characterization

X-ray diffraction (XRD) patterns of ZnO films deposited on both 182  
silicon wafer and HSR-Pt10Ir electrodes were acquired by using a 183  
Rigaku Miniflex 600 diffractometer with Cu K $\alpha$  radiation ( $\lambda = 1.5418$  184  
Å). Data were collected with a  $2\theta$  step size of  $0.02^\circ$  and a scan speed 185  
of  $1.2^\circ \text{ min}^{-1}$ . Depth profile analysis via X-ray photoelectron 186  
spectroscopy (XPS) was conducted by using a PHI VersaProbe 187  
5000 system with monochromatic Al K $\alpha$  radiation (1486.2 eV) as the 188  
X-ray source. Sputter etching was achieved using an Ar $^+$  ion gun, with 189  
10 sputtering cycles of 60 s each, followed by spectral acquisition after 190  
every cycle. The XPS spot size was set to 200  $\mu\text{m}$ , and calibration was 191  
based on the C–C component of the C 1s peak at 285.0 eV. Spectra 192  
were analyzed using Casa XPS software. Morphological analysis was 193  
carried out using a Zeiss SUPRA50VP scanning electron microscope 194  
(SEM) equipped with an in-lens Everhart-Thornley secondary 195  
electron detector. SEM micrographs of ZnO-coated HSR-Pt10Ir 196  
electrodes were captured at an accelerating voltage of 3 keV and a 197  
working distance of 5 mm. To assess the film thickness, a ZnO-coated 198  
silicon substrate was cleaved, and cross-sectional imaging was 199  
performed using SEM. Milling was performed using a TESCAN 200  
S8000X focused ion beam (FIB) microscope equipped with a Xe $^+$  ion 201  
plasma source. The milling of the HSR-Pt10Ir electrodes was carried 202  
out at a FIB current of 100 nA and an accelerating voltage of 30 kV. 203  
Elemental analysis and mapping were performed by using an Oxford 204  
UltiMax 40-mm $^2$  energy dispersive spectrometer (EDS) coupled with 205  
Aztec v3.3 software. Measurements of ZnO-coated HSR-Pt10Ir 206  
electrodes were conducted at 10 keV, targeting the M $\alpha$  lines for Pt 207  
and Ir, the L $\alpha$  line for Zn, and the K $\alpha$  line for O. 208

#### 3.3. Electrochemical Measurements

Cyclic voltammetry (CV) and electrochemical impedance spectroscopy 209  
(EIS) measurements were performed using a BioLogic VP3 210  
potentiostat in a custom-designed three-electrode Teflon cell, as used 211  
in the previous report.<sup>36</sup> CV measurements were employed as the 212  
primary electrochemical characterization technique to understand the 213  
effect of sputter-deposited ZnO coatings on the electrochemical 214  
behavior of HSR-Pt10Ir electrodes, including the specific capacitance 215  
and charge storage capacity. The obtained specific capacitance and 216  
charge storage capacity values directly correlate with the ability of 217  
implantable neural interfacing electrodes to store and/or inject charge 218  
during pulsing and to efficiently deliver charge to the surrounding 219  
tissue,<sup>37</sup> although CV measurements serve as an accelerated, model 220  
characterization rather than a direct representation of in vivo 221  
stimulation conditions.<sup>38</sup> The cell setup included a Ag/AgCl reference 222  
electrode (Gamry Instruments Inc., Warminster, PA), a coiled 223  
platinum counter electrode, and the working electrode (HSR- 224  
Pt10Ir, HSR-Pt10Ir-ZnO-5, and HSR-Pt10Ir-ZnO-60). A commer- 225  
cially available phosphate-buffered saline (PBS) solution (Blood Bank 226  
Saline, Azer Scientific) was used as the electrolyte. The applied 227  
voltage was confined to a range that avoids harmful electrochemical 228  
reactions at the interface with biological tissue or nerve.<sup>38</sup> All 229  
measurements were performed within a potential range of  $-0.6$  to  $0.8$  230  
V vs Ag/AgCl at a voltage sweep rate of  $50 \text{ mV}\cdot\text{s}^{-1}$  for 1500 cycles, 231  
with all potentials reported vs the Ag/AgCl reference electrode. The 232  
electrode–electrolyte contact area of  $13.8 \text{ mm}^2$ , calculated from the 233  
footprint of the electrode exposed to the electrolyte, was used to 234  
determine the specific capacitance and charge storage capacity from 235  
the CV data. EIS measurements were performed at the open circuit 236  
potential (OCP) over a frequency range of  $0.1$ – $10^5$  Hz, using a 237  
sinusoidal excitation voltage of 10 mV ( $V_{\text{rms}}$ ), for all the working 238  
electrodes after 15, 30, and 1500 CV cycles. 239

#### 3.4. Antibacterial Measurements

*Escherichia coli* K-12 (Gram-negative) and *Staphylococcus aureus* 240  
ATCC 27660 (Gram-positive) were used as representative bacterial 241  
strains to assess bactericidal efficacy. Active bactericidal species (e.g., 242  
Zn $^{2+}$  ions) from a coated electrode can diffuse into the bacteria- 243  
containing media on top of the agar. When the bactericidal species' 244

245 release exceeds a critical threshold, bacterial growth is inhibited in the  
 246 associated media, resulting in a zone of inhibition (ZoI) of bacterial  
 247 growth around the electrode, which appears as a transparent circle on  
 248 an otherwise opaque agar plate that is covered by growing bacterial  
 249 cells outside of the ZoI. The antibacterial performance of the  
 250 electrode surfaces was investigated using a modified Kirby–Bauer ZoI  
 251 assay.<sup>39</sup> In this adapted procedure, electrodes were aseptically placed  
 252 with the coated surface oriented downward on top of solidified agar  
 253 (VWR Life Science, Agarose RA, 7.5 g/L) containing 14 g/L Nutrient  
 254 Broth (NB; Becton Dickinson, 234000) in a Petri dish. Electrodes  
 255 were incubated on the agar surface for 2 h at room temperature to  
 256 facilitate electrode–agar interactions and ion diffusion from the  
 257 electrode into agar. Subsequently, liquid samples of log-phase  
 258 bacterial cells ( $0.5\text{--}2 \times 10^6$  CFU/plate) were spread across the  
 259 agar plates and around electrodes, which were then incubated under  
 260 completely dark, aerobic conditions at 37 °C for 18 h. Specifically, the  
 261 glass door on the incubator was covered with aluminum foil to ensure  
 262 darkness and to study the bactericidal efficacy of the electrodes under  
 263 simulated implanted/dark conditions. The concentration of colony-  
 264 forming units (CFU/mL) in each experiment was determined in  
 265 parallel cultures by diluting the initial culture 10 million-fold (i.e.,  $D =$   
 266  $10^7$ ) in NB and then spreading 1 mL of those cells onto agar plates,  
 267 which were incubated overnight at 37 °C for 18 h alongside the plates  
 268 with the electrodes. The number of colonies on each plate ( $N$ ) were  
 269 then counted, allowing us to calculate the CFU concentration of the  
 270 initial culture using eq 1.

$$271 \quad \text{CFU/mL} = D^*N/1\text{mL} \quad (1)$$

272 Following incubation, the antimicrobial efficacy of each coating was  
 273 quantified by digitally measuring the width of the ZoI based on the  
 274 corresponding electrode disc's known diameter. Due to the small  
 275 deviations in the ZoI width between each experiment, three  
 276 independent biological replicates were conducted for each type of  
 277 electrode.

#### 4. RESULTS AND DISCUSSION

278 The XRD pattern of HSR-Pt10Ir-ZnO-60 (Figure 2) shows  
 279 distinct characteristic peaks of the crystalline ZnO phase. The

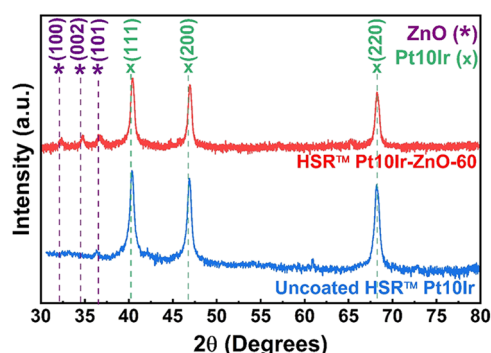


Figure 2. XRD patterns of uncoated HSR-Pt10Ir (blue-bottom) and HSR-Pt10Ir-ZnO-60 (red-top) electrodes. ZnO peaks are marked with purple dashed lines and an (\*) symbol, and Pt10Ir peaks are indicated by green dashed lines and an (x) symbol.

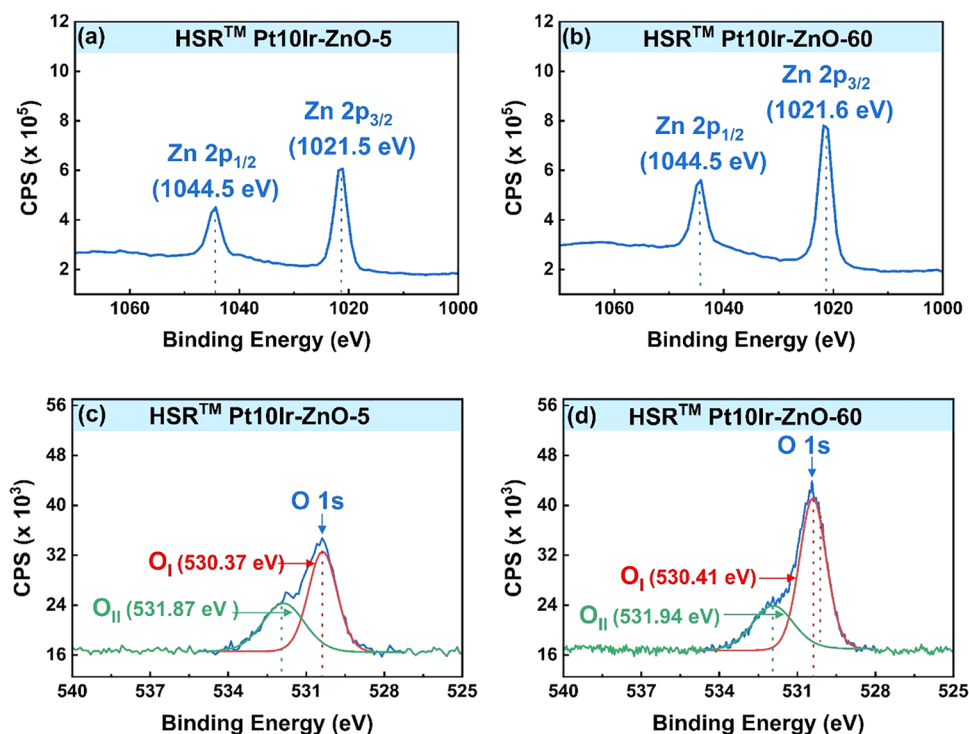
280 major peaks observed at 32.07°, 34.47°, and 36.53° values of  
 281  $2\theta$  correspond to the (100), (002), and (101) crystallographic  
 282 planes of ZnO, respectively. The grazing incidence XRD  
 283 pattern of silicon substrate coated with ZnO for 60 min  
 284 deposition (Figure S1 in the Supporting Information) shows  
 285 major peaks at 34.8 and 64.5° values of  $2\theta$  corresponding to  
 286 the (002) and (103) crystallographic planes of ZnO,  
 287 respectively. All the ZnO-coated substrates exhibit diffraction  
 288 peaks only corresponding to ZnO, indicating the absence of  
 289 secondary Zn-based phases such as metallic Zn. The substrates

coated for 5 min did not exhibit the ZnO peak, likely due to 290  
 insufficient film thickness. Cross-sectional SEM micrographs of 291  
 ZnO film deposited on a silicon substrate for 1 h (Figure S2 in 292  
 Supporting Information) revealed a thickness of approximately 293  
 280–300 nm, corresponding to a film growth rate of  $\sim 4.5$  nm· 294  
 $\text{min}^{-1}$ . Based on this rate, the thickness of ZnO films deposited 295  
 on silicon for 5 min is estimated to be around 25–30 nm. 296

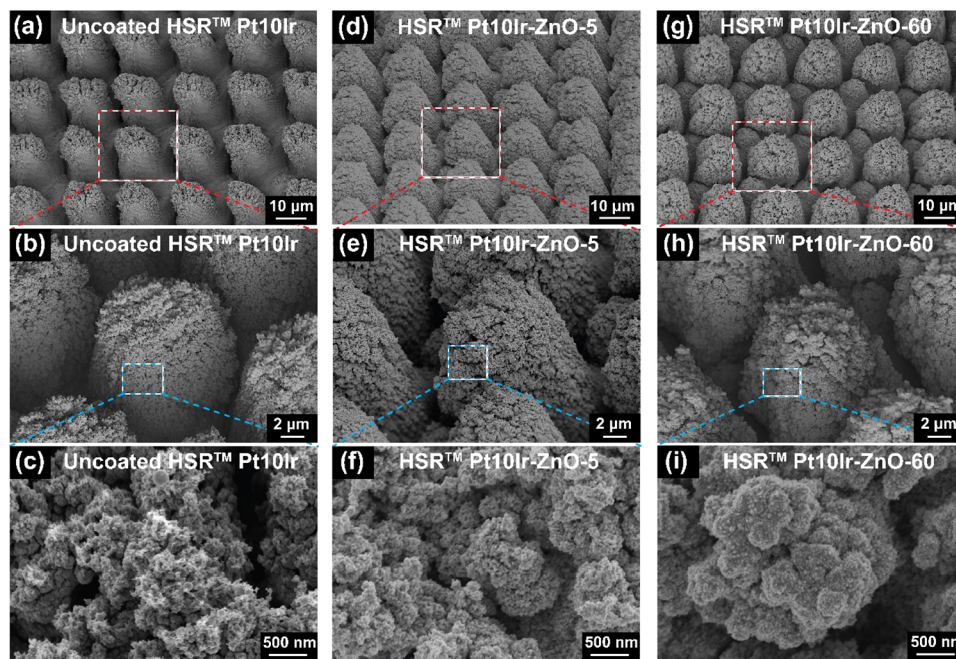
To further confirm the presence of ZnO and better 297  
 understand the characteristics of the sputtered ZnO films, 298  
 XPS analysis was performed. Figure 3 presents the XPS spectra 299 f3  
 of the ZnO-coated HSR-Pt10Ir electrodes. The survey scans of 300  
 the ZnO-coated HSR-Pt10Ir electrodes revealed characteristic 301  
 peaks corresponding to Zn 2p and O 1s, confirming the 302  
 formation of ZnO. For both HSR-Pt10Ir-ZnO-5 (Figure 3a) 303  
 and HSR-Pt10Ir-ZnO-60 (Figure 3b), the Zn 2p peak splits 304  
 into Zn 2p<sub>3/2</sub> and Zn 2p<sub>1/2</sub> due to spin–orbit interaction, with 305  
 a doublet peak energy separation of approximately 23.0 eV. In 306  
 case of both HSR-Pt10Ir-ZnO-5 and HSR-Pt10Ir-ZnO-60, the 307  
 Zn 2p<sub>3/2</sub> peak was observed at 1021.6 eV and the Zn 2p<sub>1/2</sub> 308  
 peak at 1044.5 eV. These values are well within the typical 309  
 range of binding energies reported for ZnO.<sup>40–42</sup> Figure 3c,d 310  
 present the O 1s spectra for HSR-Pt10Ir-ZnO-5 and HSR- 311  
 Pt10Ir-ZnO-60, respectively. For HSR-Pt10Ir-ZnO-5 (Figure 312  
 3c), the deconvoluted peaks were identified at 530.37 and 313  
 531.87 eV, corresponding to O(I) and O(II), respectively. The 314  
 lower binding energy peak O(I), centered at 530.37 eV, is 315  
 attributed to lattice oxygen,<sup>43,44</sup> which contributes to the 316  
 hexagonal wurtzite structure of the ZnO lattice. The higher 317  
 binding energy peak O(II), centered at 531.87 eV, has recently 318  
 been attributed to oxygen from chemisorbed water mole- 319  
 cules.<sup>44</sup> For HSR-Pt10Ir-ZnO-60 (Figure 3d), the deconvoluted 320  
 peaks of the O 1s peaks were observed at 530.41 eV for 321  
 O(I) and 531.94 eV for O(II). 322

Figure 4a–c shows SEM micrographs of uncoated HSR- 323 f4  
 Pt10Ir electrodes at increasing magnifications. The surface of 324  
 the uncoated HSR-Pt10Ir electrode exhibits a hierarchical 325  
 topography composed of periodic mound-like pillars with 326  
 diameters of  $\sim 20\text{--}25$   $\mu\text{m}$  and a valley depth of 25 to 30  $\mu\text{m}$ . 327  
 These pillars are covered by a finer nanoscale structure, with 328  
 features on the order of a few hundred nanometers in size that 329  
 contribute to the overall surface roughness. This roughness 330  
 originates from femtosecond laser restructuring and can be 331  
 tuned by varying specific laser parameters, such as average 332  
 power and fluence.<sup>8,10</sup> Figure 4d–f shows SEM micrographs of 333  
 HSR-Pt10Ir-ZnO-5 electrodes, while Figure 4g–i displays 334  
 SEM micrographs of HSR-Pt10Ir-ZnO-60 electrodes. Compar- 335  
 ison of the low-magnification SEM micrographs of uncoated 336  
 electrodes with those of sputter-coated electrodes (top row of 337  
 Figure 4) reveals that the PVD coating process does not 338  
 significantly alter the overall hierarchical surface morphology. 339  
 At higher magnifications, the surface of the uncoated HSR- 340  
 Pt10Ir pillars (Figure 4c) exhibits secondary features such as 341  
 pores and surface roughness. After ZnO deposition, as 342  
 observed in Figure 4f,i, the coating appears as agglomerated 343  
 nanospheres with diameters ranging from 40 to 60 nm. 344

Figure 5 shows the cyclic voltammograms of the electro- 345 f5  
 chemical cells containing HSR-Pt10Ir, HSR-Pt10Ir-ZnO-5, and 346  
 HSR-Pt10Ir-ZnO-60 electrodes, corresponding to the second, 347  
 15th, 30th, and 1500th cycles. The CV curves of HSR-Pt10Ir- 348  
 ZnO-5 (Figure 5b) and HSR-Pt10Ir-ZnO-60 (Figure 5c) 349  
 electrodes exhibit semirectangular shapes similar to the 350  
 uncoated HSR-Pt10Ir electrode (Figure 5a), indicating that the 351  
 capacitive behavior is retained after ZnO deposition. 352



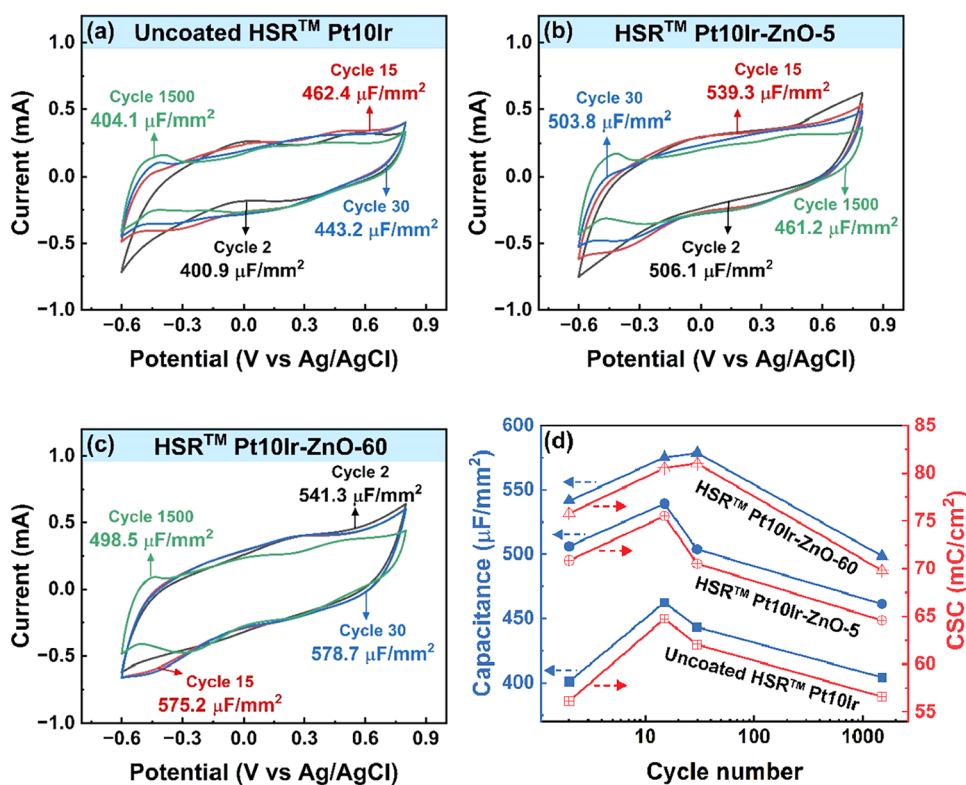
**Figure 3.** XPS spectra of HSR-Pt10Ir-ZnO-5 for (a) zinc 2p, (c) oxygen 1s, and HSR-Pt10Ir-ZnO-60 for (b) zinc 2p, (d) oxygen 1s. Deconvoluted peaks for the oxygen spectra are shown with red and green lines.



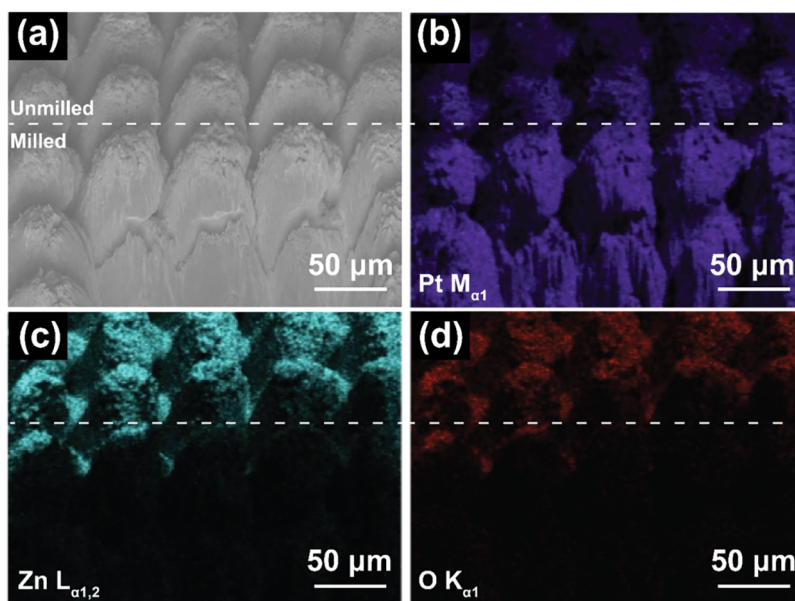
**Figure 4.** SEM micrographs of uncoated HSR-Pt10Ir (a–c), HSR-Pt10Ir-ZnO-5 (d–f), and HSR-Pt10Ir-ZnO-60 (g–i) electrodes. Micrographs (a, d, g) display the micropillar array architecture of the electrodes, micrographs (b, e, h) focus on the morphology of a single pillar, and the higher magnification micrographs (c, f, i) reveal the nanostructured morphology of the ZnO coating over the micropillars.

353 However, the area under the CV curves increased notably  
 354 following ZnO deposition. As a result, the specific capacitance,  
 355 calculated from the second CV cycle, increased by  $105 \mu\text{F}\cdot\text{mm}^{-2}$   
 356 for HSR-Pt10Ir-ZnO-5 ( $506.1 \mu\text{F}\cdot\text{mm}^{-2}$ ) and  $140$   
 357  $\mu\text{F}\cdot\text{mm}^{-2}$  for HSR-Pt10Ir-ZnO-60 ( $541.3 \mu\text{F}\cdot\text{mm}^{-2}$ ) electro-  
 358 des compared to the uncoated HSR-Pt10Ir ( $400.9 \mu\text{F}\cdot\text{mm}^{-2}$ )  
 359 electrode. This increase in the specific capacitance confirms the

presence of a new material on the coated electrode surface. 360  
 Despite the assumption that ZnO's moderately conductive 361  
 nature<sup>45</sup> might diminish the performance of HSR-Pt10Ir 362  
 electrodes, the specific capacitance of both HSR-Pt10Ir-ZnO-5 363  
 and HSR-Pt10Ir-ZnO-60 electrodes was higher than that of 364  
 uncoated ones. To understand this enhancement, the 365  
 conformance of sputtered ZnO film over the HSR topography 366



**Figure 5.** Cyclic voltammograms of (a) uncoated HSR-Pt10Ir, (b) HSR-Pt10Ir-ZnO-5, and (c) HSR-Pt10Ir-ZnO-60 electrodes measured at a scan rate of  $50 \text{ mV}\cdot\text{s}^{-1}$ . Specific capacitance values at each cycle are indicated on the respective plots. (d) Evolution of specific capacitance (left y-axis) and charge storage capacity (right y-axis) as a function of CV cycle number (log scale) for all three electrodes: uncoated HSR-Pt10Ir (squares), HSR-Pt10Ir-ZnO-5 (circles), and HSR-Pt10Ir-ZnO-60 (triangles).



**Figure 6.** SEM micrographs and elemental EDS mapping of the HSR-Pt10Ir-ZnO-5 surface after FIB cross-sectioning. (a) SEM micrograph; (b) EDS mapping of platinum; (c) EDS mapping of zinc; and (d) EDS mapping of oxygen.

367 was examined, as capacitance is largely governed by the  
368 material's electrochemically active surface area (ESA).

369 The conformality of the sputtered ZnO films was examined  
370 by using FIB cross-sectioning and EDS mapping, as shown in  
371 Figure 6. Selected pillars from the HSR-Pt10Ir-ZnO-5  
372 electrode were milled to expose the lateral surfaces of the  
373 adjacent pillars. Subsequent EDS mapping of the area (Figure

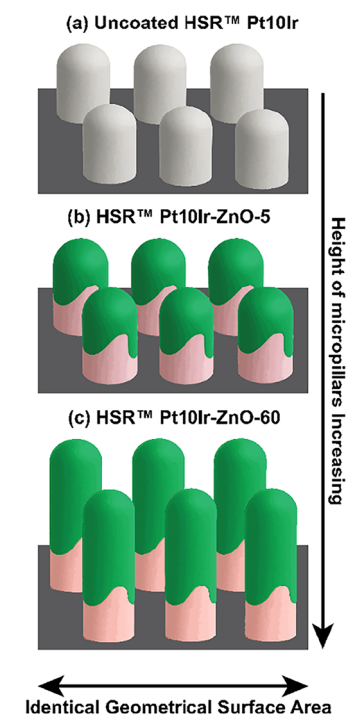
6c,d) confirmed the presence of zinc and oxygen on the HSR-  
Pt10Ir surface. The analysis revealed that while the tops of the  
pillars were predominantly coated with ZnO, the sides  
exhibited only minimal coverage. This indicates that the  
sputtered ZnO films lack conformality and do not evenly coat  
the pillar-like structures. Instead, ZnO primarily increases the  
height of the pillars without providing uniform coverage. This

381 uneven film distribution effectively increases the active ESA  
382 within the same footprint, contributing to higher capacitance  
383 values.

384 Additionally, the partial ZnO coating leaves some regions of  
385 the Pt10Ir pillars exposed and uncoated. This observation  
386 prompted further investigation of the nature of these uncoated  
387 areas. Since the ZnO deposition occurs in an oxidizing  
388 environment inside the sputtering chamber, there is a  
389 possibility that the Ir in the HSR-Pt10Ir substrate may oxidize,  
390 given the high affinity of Ir toward oxidation. A mock  
391 sputtering experiment was conducted to investigate this  
392 phenomenon. The HSR Pt10Ir substrate was placed inside  
393 the sputtering chamber under the same conditions used for  
394 ZnO deposition, except that the substrate stage was oriented  
395 away from the Zn target to prevent any ZnO coating. After 5  
396 min, the substrate (now referred to as oxidized HSR-Pt10Ir)  
397 was removed, and depth-profile XPS analysis was performed to  
398 determine whether Ir oxidation had occurred and penetrated  
399 beneath the surface. The CV results and XPS spectra of Ir 4f  
400 after 2 min of surface etching obtained from oxidized HSR-  
401 Pt10Ir electrode are shown in Figure S3a,b in the Supporting  
402 Information, respectively. Following peak fitting,<sup>46</sup> the binding  
403 energies and relative area percentages of the deconvoluted  
404 peaks are summarized in Table S1 in Supporting Information.  
405 An increase in the relative area associated with the Ir<sup>4+</sup> species  
406 was observed after the mock oxidation experiment. Hence, it  
407 was confirmed that the exposed uncoated regions of the ZnO-  
408 coated HSR-Pt10Ir contain oxidized Ir. Due to the non-  
409 conformal nature of the sputtered ZnO coating, certain parts of  
410 the HSR surface remain exposed without ZnO coating,  
411 allowing the partially oxidized Pt10Ir substrate to be in  
412 contact with the electrolyte. The presence of exposed  
413 conductive regions supports efficient electron transport despite  
414 the moderate conductivity of the ZnO coating. However,  
415 surface Ir oxidation is not considered to be the primary origin  
416 of the observed capacitance enhancement. While oxidized Ir in  
417 the partially exposed Pt10Ir metallic surface of the electrode  
418 may provide a secondary contribution to the electrochemical  
419 response, the dominant mechanism governing the increased  
420 specific capacitance of the HSR-Pt10Ir-ZnO-5 and HSR-  
421 Pt10Ir-ZnO-60 electrodes is likely the increase in electro-  
422 chemically active surface area resulting from the nonconformal  
423 ZnO coating.

424 A schematic illustration of the nonconformal growth of  
425 sputtered ZnO films on the HSR-Pt10Ir electrode surface is  
426 presented in Figure 7. As shown in Figure 7b,c, ZnO  
427 deposition increases the height of the micropillars with longer  
428 deposition times, leading to a higher ESA within the same  
429 geometric surface area. Due to the nonconformal nature of the  
430 ZnO deposition, the regions along the micropillar sidewalls  
431 remain uncoated, leaving the underlying Pt10Ir surface  
432 uncoated and, thus, partially oxidized.

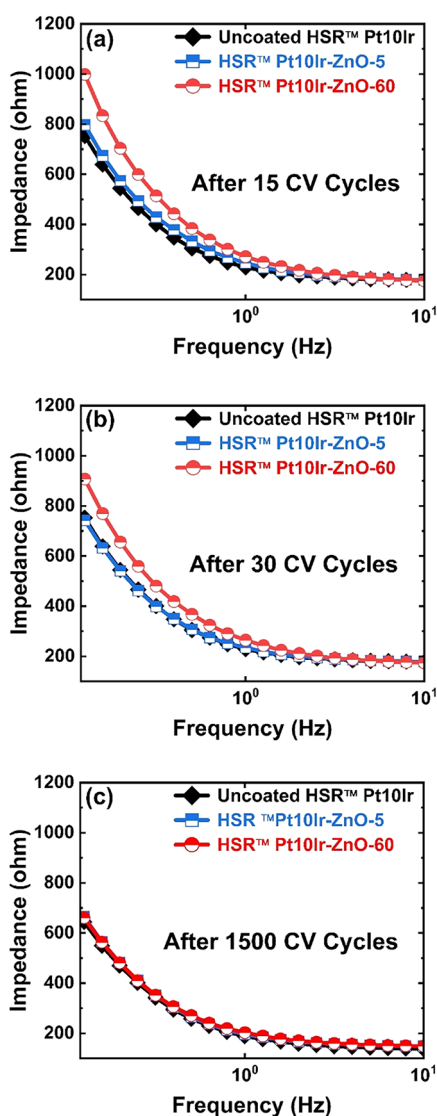
433 Electrochemical impedance spectroscopy (EIS) was used to  
434 investigate the evolution of the HSR-Pt10Ir-ZnO-5 and HSR-  
435 Pt10Ir-ZnO-60 electrodes during prolonged cycling. Bode  
436 impedance plots of the HSR-Pt10Ir, HSR-Pt10Ir-ZnO-5, and  
437 HSR-Pt10Ir-ZnO-60 electrodes after 15, 30, and 1500 CV  
438 cycles are shown in Figure 8a–8c, respectively. The analysis  
439 indicates that after 15 CV cycles (Figure 8a), the impedance of  
440 the HSR-Pt10Ir-ZnO-5 and HSR-Pt10Ir-ZnO-60 electrodes is  
441 notably higher than that of the uncoated HSR-Pt10Ir  
442 electrode. The increased impedance observed in HSR-Pt10Ir-  
443 ZnO-5 and HSR-Pt10Ir-ZnO-60 electrodes during initial



**Figure 7.** Schematic illustration showing the growth and nonuniform coverage of the ZnO coating (green) on HSR-Pt10Ir electrodes along with Ir oxidation (pink) induced during sputtering: (a) uncoated HSR Pt10Ir, (b) HSR-Pt10Ir-ZnO-5, and (c) HSR-Pt10Ir-ZnO-60 electrodes. The change in the pillar height is shown for illustration purposes only and is not drawn to scale.

444 cycling is attributed to the ZnO coating, which introduces  
445 additional resistance to electron transport due to its lower  
446 conductivity compared with the metallic Pt10Ir. However, after  
447 30 cycles (Figure 8b), the impedance of the HSR-Pt10Ir-ZnO-  
448 5 electrode decreases and approaches the value of the uncoated  
449 HSR-Pt10Ir electrode. Similarly, after 1500 cycles (Figure 8c),  
450 the impedances of both HSR-Pt10Ir-ZnO-5 and HSR-Pt10Ir-  
451 ZnO-60 electrodes decrease and match the value of the  
452 uncoated electrode. These observations suggest a gradual  
453 removal of the ZnO layer, likely due to the dissolution in  
454 electrolyte, which exposes more of the conductive Pt10Ir  
455 surface and reduces the overall resistivity, leading to improved  
456 electron transport and lower impedance as observed in the EIS  
457 Bode plots.

458 The gradual stripping of ZnO during CV cycling was further  
459 investigated using postcycling SEM and EDS analyses. Figure 9  
460 shows the postcycling SEM micrographs and EDS mapping of  
461 HSR-Pt10Ir-ZnO-5 and HSR-Pt10Ir-ZnO-60 electrodes. The  
462 cycling was stopped after 1500 CV cycles (approximately 24 h  
463 of electrochemical cycling) at  $-0.6$  V. Post-CV EDS analysis  
464 confirms that following 1500 CV cycles, the sputtered ZnO  
465 layer is no longer detectable on the electrode surface for either  
466 HSR-Pt10Ir-ZnO-5 (Figure 9a–d) or HSR-Pt10Ir-ZnO-60  
467 (Figure 9e–h) electrodes. This observation indicates that,  
468 under the applied potential window used during continuous  
469 CV cycling, the sputtered ZnO coating progressively dissolves  
470 into the electrolyte, a process that is integral to the  
471 antibacterial functionality of ZnO in the surrounding medium.  
472 ZnO is widely reported<sup>23,28</sup> to exhibit antibacterial activity  
473 through the release and diffusion of Zn<sup>2+</sup> ions into the  
474 environment, where these ions can disrupt bacterial cell walls



**Figure 8.** Bode impedance plots of uncoated HSR-Pt10Ir (black), HSR-Pt10Ir-ZnO-5 (blue), and HSR-Pt10Ir-ZnO-60 (red), measured after (a) 15, (b) 30, and (c) 1500 CV cycles.

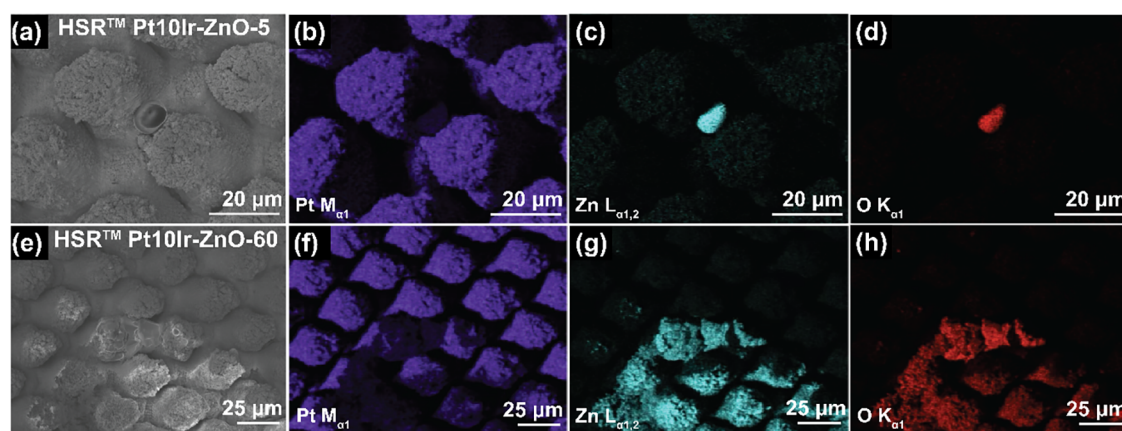
475 and interfere with intracellular processes, ultimately resulting in  
476 bactericidal effects.

The ZnO dissolution behavior observed during cyclic 477  
478 voltammetry is governed by the electrochemical conditions  
479 imposed by continuous cycling across a wide potential  
480 window. In practical device operation, implantable electrodes  
481 are not subjected to such sustained polarization but instead  
482 operate under low-amplitude, charge-balanced pulsed voltage,  
483 or current stimulation. Under these clinically relevant regimes,  
484 the electrochemical driving forces that govern ZnO dissolution  
485 are expected to differ substantially, resulting in a dissolution  
486 behavior that more closely reflects long-term in vivo  
487 conditions.

In addition to ZnO dissolution, as a consequence of the 488  
489 confined electrolyte volume during CV measurements, localized Zn- and O-rich features (Figure 9c–g,h) were also  
490 observed after 1500 CV cycles, indicating ZnO recrystallization  
491 at isolated surface sites. While ZnO recrystallization is typically  
492 reported at more negative potentials ( $-0.8$  to  $-2$  V vs Ag/  
493 AgCl),<sup>47–49</sup> the hierarchical surface geometry of the HSR-  
494 Pt10Ir electrodes may give rise to spatially heterogeneous  
495 electrochemical environments,<sup>50–52</sup> which could facilitate  
496 limited, site-specific redeposition even at  $-0.6$  V of applied  
497 potential.  
498

The observed ZnO recrystallization also serves as indirect 499  
500 evidence of  $Zn^{2+}$  ion release into the electrolyte, which is a  
501 well-established antibacterial mechanism for ZnO. Further-  
502 more, the occurrence of localized recrystallization likely  
503 suggests that, under the specific CV conditions, the local  
504 electrolyte environment became enriched with  $Zn^{2+}$  ions,  
505 indicating a high amount of ZnO mass loading on the HSR-  
506 Pt10Ir electrode surface that can support its antibacterial  
507 functionality. During device-specific stimulation pulses, the  
508 dissolution of  $Zn^{2+}$  ions is expected to proceed at a different  
509 rate, and the released  $Zn^{2+}$  ions into the surrounding  
510 environment will likely be consumed through antibacterial  
511 interactions or diffuse away from the electrode surface into the  
512 bloodstream. Under such conditions, the accumulation of Zn  
513 species required for recrystallization is unlikely, and ZnO  
514 dissolution will likely remain the dominant and functionally  
515 relevant process.

Based on the CV cycling and postcycling characterization of 516  
517 HSR-Pt10Ir-ZnO electrodes presented above, we propose the  
518 following mechanism for the evolution of ZnO coatings during  
519 prolonged electrochemical cycling: when exposed to the saline  
520 solution electrolyte during CV cycling, HSR-Pt10Ir-ZnO-5 and



**Figure 9.** SEM micrographs and EDS mapping of (a)–(d) HSR-Pt10Ir-ZnO-5 electrode, and (e)–(h) HSR-Pt10Ir-ZnO-60 electrode, after 1500 CV cycles (24 h extended cycling) stopped at  $-0.6$  V.

521 HSR-Pt10Ir-ZnO-60 electrodes exhibit higher capacitances  
522 than the uncoated HSR-Pt10Ir electrode, primarily due to the  
523 increased ESA resulting from the sputtered, nonconformal  
524 ZnO layer. Additionally, the increased capacitance can be  
525 attributed to the partially oxidized Ir in the composition of the  
526 exposed uncoated regions of the HSR-Pt10Ir electrode.  
527 Despite the moderate conductivity of the ZnO film, the  
528 overall electron transport remains efficient, supported by the  
529 underlying and uncoated metallic Pt10Ir regions of the  
530 electrode surface. As cycling progresses, the deposited ZnO  
531 layer gradually strips from the electrode surface and dissolves  
532 into the electrolyte, consistent with the observed decrease in  
533 impedance of the ZnO-coated electrodes over extended  
534 electrochemical cycling. ZnO dissolution was further con-  
535 firmed through postcycling EDS mapping of the ZnO-coated  
536 HSR-Pt10Ir electrodes. After extended cycling, SEM micro-  
537 graphs and EDS mapping revealed localized recrystallization of  
538 ZnO on the electrode surface, forming well-faceted crystals.  
539 The agglomeration of bulky recrystallized ZnO particles  
540 reduces the available ESA, leading to a decrease in the specific  
541 capacitance at later cycles (i.e., cycle 1500). Notably, the HSR-  
542 Pt10Ir-ZnO-60 electrode, with a thicker ZnO layer, experi-  
543 ences prolonged ZnO dissolution that delays recrystallization,  
544 thereby better preserving the capacitance during extended  
545 cycling.

546 While the electrochemical measurements in this study were  
547 carried out in a confined environment with continuous voltage  
548 application during CV, which allowed the recrystallization of  
549 dissolved ZnO on the electrode surface, in practical  
550 implantable applications, the electrodes will be subjected to  
551 pulsed currents or voltages within a larger biological environ-  
552 ment. Under these conditions, the dissolved  $Zn^{2+}$  ions are  
553 expected to interact with and diffuse into the bacterial cell walls  
554 to eliminate them.<sup>28</sup> This leads to subsequent consumption of  
555  $Zn^{2+}$  ions, therefore, preventing ZnO recrystallization on the  
556 electrode surface.

557 To verify the antibacterial activity of the electrodes, a  
558 modified Kirby–Bauer zone of inhibition (ZOI) method was  
559 used to test the ZnO-coated HSR-Pt10Ir electrodes against *S.*  
560 *aureus* (Gram-positive) and *E. coli* (Gram-negative) bacteria.  
561 The HSR-Pt10Ir-ZnO-60 electrode was selected for this study  
562 as it has a higher ZnO mass loading and demonstrated the best  
563 electrochemical performance. Figure 10 shows that the HSR-  
564 Pt10Ir-ZnO-60 electrode exhibits a clear ZOI as compared to  
565 uncoated HSR-Pt10Ir electrodes against both *S. aureus* (Figure  
566 10a) and *E. coli* (Figure 10b) bacterial strains. The average ZOI  
567 width observed for *E. coli* was  $3.93 \pm 0.46$  mm, while the ZOI  
568 for *S. aureus* was  $2.10 \pm 0.42$  mm, indicating that ZnO has  
569 higher bactericidal efficacy for *E. coli* compared to *S. aureus*.

570 The inhibition of bacterial growth extending beyond the  
571 ZnO-coated surfaces indicates that antimicrobial species  
572 diffuse from the coating into the surrounding medium. As all  
573 experiments were conducted under dark conditions, the active  
574 species are most likely  $Zn^{2+}$  ions released from the ZnO layer.  
575 This ion release appears sufficient to achieve effective  
576 inhibition of both bacterial strains, likely through  $Zn^{2+}$ -  
577 mediated disruption of cell membrane integrity and associated  
578 impairment of cellular function. These findings demonstrate  
579 that sputter-deposited ZnO coatings can reliably endow HSR-  
580 Pt10Ir electrodes with antibacterial functionality.

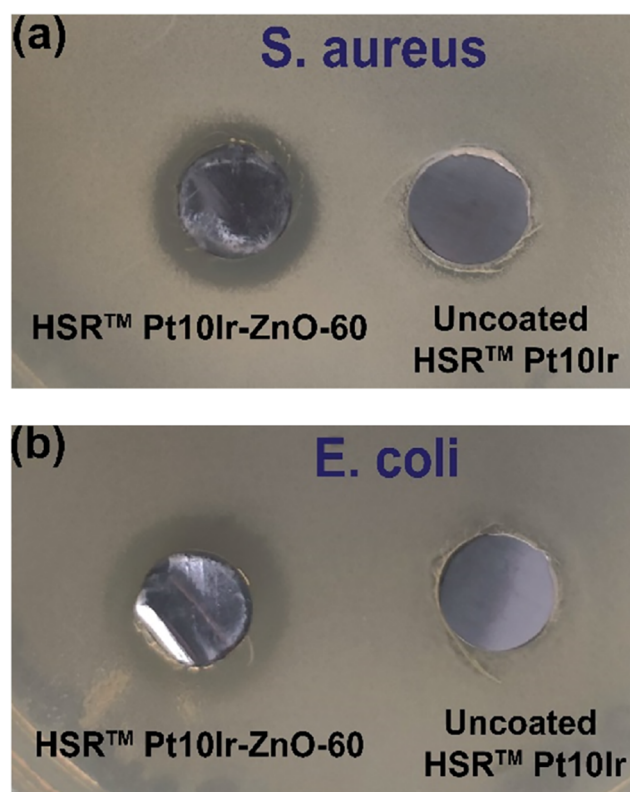


Figure 10. Antibacterial activity of HSR-Pt10Ir-ZnO-60 electrode alongside uncoated HSR-Pt10Ir electrode against (a) *S. aureus* and (b) *E. coli* bacterial strains. ZOI is clearly visible for HSR-Pt10Ir-ZnO-60 electrodes as a ring of different contrast around them. The electrodes are 10 mm discs, which can be used as scale bars to measure ZOI.

## 5. CONCLUDING REMARKS

This study establishes a dual-functional electrode platform in which reactively sputtered zinc oxide (ZnO) coatings, integrated with hierarchically restructured (HSR) platinum–iridium (Pt10Ir) electrodes, synergistically enhance electrochemical performance while imparting robust antibacterial functionality. Deposition of ZnO across the hierarchically restructured Pt10Ir electrode surface increases the electrochemically active surface area (ESA), resulting in specific capacitance enhancements of approximately 16% and 24% for the HSR-Pt10Ir-ZnO-5 (at ~30 nm ZnO) and HSR-Pt10Ir-ZnO-60 (at ~300 nm ZnO) electrodes, respectively, relative to uncoated HSR-Pt10Ir control electrodes. X-ray photoelectron spectroscopy (XPS) confirms the formation of oxidized Ir species within partially exposed micropillar regions, indicating that controlled surface oxidation facilitates efficient charge transport and partially contributes to the observed capacitance enhancement.

Electrochemical impedance spectroscopy (EIS) reveals a progressive decrease in the interfacial impedance during extended cycling, consistent with gradual ZnO dissolution into the electrolyte. Following prolonged cyclic voltammetry (CV), localized ZnO recrystallization is also observed on the electrode surface. However, continuous CV cycling over wide potential windows does not reflect the practical device operation. Implantable neurostimulation and cardiac rhythm management electrodes are engineered for long-term stimulation using low-amplitude, charge-balanced, constant-current

608 pulsed waveforms, under which the electrochemical conditions  
609 that promote ZnO dissolution and recrystallization during  
610 extended CV are unlikely to arise. The dissolution and stability  
611 behavior observed under prolonged CV cycling should  
612 therefore be interpreted as an accelerated electrochemical  
613 stress condition rather than a direct analogue of in vivo  
614 performance.

615 Taken together, the results of this study provide a  
616 foundational demonstration of integrating sputter-deposited,  
617 cost-effective, and commercially viable ZnO coatings with  
618 HSR-Pt10Ir electrodes. The observed antibacterial activity  
619 confirms that ZnO-coated HSR-Pt10Ir electrodes can  
620 effectively suppress bacterial proliferation while preserving  
621 and, in some cases, enhancing electrochemical performance.  
622 Collectively, these findings identify sputter-deposited ZnO as a  
623 promising materials integration strategy for implantable  
624 neurostimulation and cardiac rhythm management applica-  
625 tions.

626 Future work will focus on long-term stimulation studies  
627 under physiologically relevant conditions using pulsed  
628 electrical signals and simulated body fluids to further assess  
629 coating stability, sustained antibacterial activity, and overall  
630 electrode performance.

## 631 ■ ASSOCIATED CONTENT

### 632 ■ Supporting Information

633 The Supporting Information is available free of charge at  
634 <https://pubs.acs.org/doi/10.1021/acsami.5c21727>.

635 XRD pattern of Si wafer coated with ZnO for a 60 min  
636 deposition duration, cross-sectional SEM micrograph of  
637 cleaved ZnO-coated planar Si wafer deposited for 60  
638 min, CV cycling results of oxidized HSR-Pt10Ir  
639 electrode, XPS spectra, and peak fitting analysis of  
640 oxidized HSR-Pt10Ir electrode (PDF)

## 641 ■ AUTHOR INFORMATION

### 642 Corresponding Authors

643 Ekaterina Pomerantseva – Department of Materials Science  
644 and Engineering, Drexel University, Philadelphia,  
645 Pennsylvania 19104, United States; [orcid.org/0000-0002-6765-7133](https://orcid.org/0000-0002-6765-7133); Email: [ep423@drexel.edu](mailto:ep423@drexel.edu)

647 Shahram Amini – Pulse Technologies Inc. (An Integer  
648 Holdings Company), Quakertown, Pennsylvania 18951,  
649 United States; Biomedical Engineering Department,  
650 University of Connecticut, Storrs, Connecticut 06269, United  
651 States; Email: [shahram.amini@integer.net](mailto:shahram.amini@integer.net)

### 652 Authors

653 Kriti Panchal – Department of Materials Science and  
654 Engineering, Drexel University, Philadelphia, Pennsylvania  
655 19104, United States; [orcid.org/0009-0002-9778-7104](https://orcid.org/0009-0002-9778-7104)

656 Wesley Seche – Pulse Technologies Inc. (An Integer Holdings  
657 Company), Quakertown, Pennsylvania 18951, United  
658 States; [orcid.org/0009-0008-5842-4152](https://orcid.org/0009-0008-5842-4152)

659 Henna Khosla – Department of Mechanical Engineering,  
660 Villanova University, Villanova, Pennsylvania 19085, United  
661 States; [orcid.org/0000-0003-3136-9276](https://orcid.org/0000-0003-3136-9276)

662 Gang Feng – Department of Mechanical Engineering,  
663 Villanova University, Villanova, Pennsylvania 19085, United  
664 States; [orcid.org/0000-0002-4952-3772](https://orcid.org/0000-0002-4952-3772)

Jacob Elmer – Department of Chemical and Biological  
Engineering, Villanova University, Villanova, Pennsylvania  
19085, United States; [orcid.org/0000-0001-6980-489X](https://orcid.org/0000-0001-6980-489X)

Gregory A. Caputo – Department of Chemistry and  
Biochemistry, Rowan University, Glassboro, New Jersey  
08028, United States; [orcid.org/0000-0002-4510-2815](https://orcid.org/0000-0002-4510-2815)

Steven J. May – Department of Materials Science and  
Engineering, Drexel University, Philadelphia, Pennsylvania  
19104, United States; [orcid.org/0000-0002-8097-1549](https://orcid.org/0000-0002-8097-1549)

Complete contact information is available at:  
<https://pubs.acs.org/10.1021/acsami.5c21727>

## 676 Notes

The authors declare no competing financial interest. 677

## 678 ■ ACKNOWLEDGMENTS

The authors gratefully acknowledge the Materials Character-  
ization Core (MCC) at Drexel University for providing access  
to advanced materials characterization facilities. A subset of  
this work was carried out at the Singh Center for Nano-  
technology at the University of Pennsylvania, which is  
supported by the National Science Foundation (NSF)  
National Nanotechnology Coordinated Infrastructure  
(NNCI) Program under Grant No. NNCI-2025608. The  
authors extend special thanks to Dmitri Barbash for training  
and technical support on MCC instrumentation and to Prof.  
Jeff Hettinger of Rowan University for expert guidance on FIB-  
SEM analysis. This work was supported in part by the  
Manufacturing PA Innovation Program of the Pennsylvania  
Department of Community and Economic Development.  
Primary financial support from Joseph Rosato, Frank Henofer,  
and Bob Walsh of Pulse IP, LLC is sincerely appreciated for  
their continued commitment to advancing innovative medical  
device technologies.

## 697 ■ REFERENCES

- (1) Epstein, L. J.; Palmieri, M. Managing chronic pain with spinal cord stimulation. *Mount Sinai J. Med.* **2012**, *79* (1), 123–132. 698
- (2) Ordonez, J. S.; Boehler, C.; Schuettler, M.; Stieglitz, T. Improved Polyimide Thin-Film Electrodes for Neural Implants. In *2012 Annual International Conference of the IEEE Engineering in Medicine and Biology Society*; IEEE, 2012; Vol. 2012, pp 5134–5137. 699
- (3) Stenehjelm, E.; Armstrong, W. S. Central Nervous System Device Infections. *Infect. Dis. Clin. North Am.* **2012**, *26* (1), 89–110. 700
- (4) Schalk, G.; Leuthardt, E. C. Brain-Computer Interfaces Using Electrocorticographic Signals. *IEEE Rev. Biomed. Eng.* **2011**, *4*, 140–154. 701
- (5) Mulpuru, S. K.; Madhavan, M.; McLeod Christopher, J.; Cha, Y.-M.; Friedman Paul, A. Cardiac Pacemakers: Function, Troubleshooting, and Management. *J. Am. Coll. Cardiol.* **2017**, *69* (2), 189–210. 702
- (6) Stevenson, I.; Voskoboinik, A. Cardiac rhythm management devices. *Aust. J. Gen. Pract.* **2018**, *47*, 264–271. 703
- (7) Eljamel, S.; Slavin, K. *Neurostimulation: Principles and Practice*; John Wiley & Sons, 2013. 704
- (8) Amini, S.; Seche, W.; May, N.; Choi, H.; Tavousi, P.; Shahbazmohamadi, S. Femtosecond laser hierarchical surface restructuring for next generation neural interfacing electrodes and microelectrode arrays. *Sci. Rep.* **2022**, *12* (1), No. 13966. 705
- (9) Amini, S.; Choi, H.; Seche, W.; Blagojevic, A.; May, N.; Lefler, B. M.; Davis, S. L.; Elyahoodayan, S.; Tavousi, P.; May, S. J.; et al. Sustainability inspired fabrication of next generation neurostimulation and cardiac rhythm management electrodes via reactive hierarchical surface restructuring. *Microsyst. Nanoeng.* **2024**, *10* (1), 125. 706

- 726 (10) Blagojevic, A.; Seche, W.; Choi, H.; Davis, S. L.; Elyahoodayan,  
727 S.; Caputo, G. A.; Lowe, T. C.; Tavousi, P.; Shahbazmohamadi, S.;  
728 Amini, S. Hierarchical Surface Restructuring of Ultra-Thin Electrodes  
729 and Microelectrode Arrays for Neural Interfacing with Peripheral and  
730 Central Nervous Systems. *Adv. Mater. Interfaces* **2024**, *11* (23),  
731 No. 2400017.
- 732 (11) Makino, T.; Jimi, S.; Oyama, T.; Nakano, Y.; Hamamoto, K.;  
733 Mamishin, K.; Yahiro, T.; Hara, S.; Takata, T.; Ohjimi, H. Infection  
734 mechanism of biofilm-forming *Staphylococcus aureus* on indwelling  
735 foreign materials in mice. *Int. Wound J.* **2015**, *12* (2), 122–131.
- 736 (12) Fuglestad, M. A.; Tracey, E. L.; Leinicke, J. A. Evidence-based  
737 prevention of surgical site infection. *Surg. Clin.* **2021**, *101* (6), 951–  
738 966.
- 739 (13) Razavi, B.; Rao, V. R.; Lin, C.; Bujarski, K. A.; Patra, S. E.;  
740 Burdette, D. E.; Geller, E. B.; Brown, M. G. M.; Johnson, E. A.; Drees,  
741 C.; et al. Real-world experience with direct brain-responsive  
742 neurostimulation for focal onset seizures. *Epilepsia* **2020**, *61* (8),  
743 1749–1757.
- 744 (14) Krames, E.; Peckham, P. H.; Rezai, A. R. *Neuromodulation:*  
745 *Comprehensive Textbook of Principles, Technologies, and Therapies*;  
746 Academic Press, 2018.
- 747 (15) Ban, K. A.; Minei, J. P.; Laronga, C.; Harbrecht, B. G.; Jensen,  
748 E. H.; Fry, D. E.; Itani, K. M.; Dellinger, P. E.; Ko, C. Y.; Duane, T. M.  
749 American College of Surgeons and Surgical Infection Society: surgical  
750 site infection guidelines, 2016 update. *J. Am. Coll. Surg.* **2017**, *224* (1),  
751 59–74.
- 752 (16) Hood, M. L.; Skaar, E. P. Nutritional immunity: transition  
753 metals at the pathogen–host interface. *Nat. Rev. Microbiol.* **2012**, *10*  
754 (8), 525–537.
- 755 (17) Mostafavi, E.; Dubey, A. K.; Walkowiak, B.; Kaushik, A.;  
756 Ramakrishna, S.; Teodori, L. Antimicrobial surfaces for implantable  
757 cardiovascular devices. *Curr. Opin. Biomed. Eng.* **2022**, *23*,  
758 No. 100406.
- 759 (18) Geyao, L.; Yang, D.; Wanglin, C.; Chengyong, W. Development  
760 and Application of Physical Vapor Deposited Coatings for Medical  
761 Devices: A Review. In *CIRP BioManufacturing Conference 2019*;  
762 Procedia CIRP: Guangzhou, China, 2020; Vol. 89, pp 250–262.
- 763 (19) Schmitz, T. Functional Coatings by Physical Vapor Deposition  
764 (PVD) for Biomedical Applications. Ph.D. Thesis, Universität  
765 Würzburg, 2016.
- 766 (20) Abebe, B.; Zereffa, E. A.; Tadesse, A.; Murthy, H. A. A review  
767 on enhancing the antibacterial activity of ZnO: Mechanisms and  
768 microscopic investigation. *Nanoscale Res. Lett.* **2020**, *15*, 1–19.
- 769 (21) Zhang, T.; Wang, W.; Liu, J.; Wang, L.; Tang, Y.; Wang, K. A  
770 review on magnesium alloys for biomedical applications. *Front. Bioeng.*  
771 *Biotechnol.* **2022**, *10*, No. 953344.
- 772 (22) Zhang, E.; Zhao, X.; Hu, J.; Wang, R.; Fu, S.; Qin, G.  
773 Antibacterial metals and alloys for potential biomedical implants.  
774 *Bioact. Mater.* **2021**, *6* (8), 2569–2612.
- 775 (23) Espitia, P. J. P.; Soares, N. d. F. F.; Coimbra, J. S. d. R.; de  
776 Andrade, N. J.; Cruz, R. S.; Medeiros, E. A. A. Zinc oxide  
777 nanoparticles: synthesis, antimicrobial activity and food packaging  
778 applications. *Food Bioprocess Technol.* **2012**, *5*, 1447–1464.
- 779 (24) Wang, D.; Hosteen, O.; Fierke, C. A. ZntR-mediated  
780 transcription of zntA responds to nanomolar intracellular free zinc.  
781 *J. Inorg. Biochem.* **2012**, *111*, 173–181.
- 782 (25) Jalal, R.; Goharshadi, E. K.; Abareshi, M.; Moosavi, M.; Yousefi,  
783 A.; Nancarrow, P. ZnO nanofluids: green synthesis, characterization,  
784 and antibacterial activity. *Mater. Chem. Phys.* **2010**, *121* (1–2), 198–  
785 201.
- 786 (26) Kasemets, K.; Ivask, A.; Dubourguier, H.-C.; Kahru, A. Toxicity  
787 of nanoparticles of ZnO, CuO and TiO<sub>2</sub> to yeast *Saccharomyces*  
788 *cerevisiae*. *Toxicol. Vitro* **2009**, *23* (6), 1116–1122.
- 789 (27) Zhang, L.; Ding, Y.; Povey, M.; York, D. ZnO nanofluids—A  
790 potential antibacterial agent. *Prog. Nat. Sci.* **2008**, *18* (8), 939–944.
- 791 (28) Joe, A.; Park, S.-H.; Shim, K.-D.; Kim, D.-J.; Jhee, K.-H.; Lee,  
792 H.-W.; Heo, C.-H.; Kim, H.-M.; Jang, E.-S. Antibacterial mechanism  
793 of ZnO nanoparticles under dark conditions. *J. Ind. Eng. Chem.* **2017**,  
794 *45*, 430–439.
- (29) Padmavathy, N.; Vijayaraghavan, R. Enhanced bioactivity of  
ZnO nanoparticles—an antimicrobial study. *Sci. Technol. Adv. Mater.* **2008**, *9* (3), No. 035004. 797
- (30) Puspasari, V.; Ridhova, A.; Hermawan, A.; Amal, M. I.; Khan,  
M. M. ZnO-based antimicrobial coatings for biomedical applications. *Bioprocess Biosyst. Eng.* **2022**, *45* (9), 1421–1445. 800
- (31) Lakshmi Prasanna, V.; Vijayaraghavan, R. Insight into the  
Mechanism of Antibacterial Activity of ZnO: Surface Defects  
Mediated Reactive Oxygen Species Even in the Dark. *Langmuir* **2015**, *31* (33), 9155–9162. 804
- (32) Nagvenkar, A. P.; Deokar, A.; Perelshtein, I.; Gedanken, A. A  
one-step sonochemical synthesis of stable ZnO–PVA nanocolloid as a  
potential biocidal agent. *J. Mater. Chem. B* **2016**, *4* (12), 2124–2132. 807
- (33) Lawrence, M. C.; Pilling, P. A.; Epa, V. C.; Berry, A. M.;  
Ogunniyi, A. D.; Paton, J. C. The crystal structure of pneumococcal  
surface antigen PsaA reveals a metal-binding site and a novel structure  
for a putative ABC-type binding protein. *Structure* **1998**, *6* (12),  
1553–1561. 812
- (34) Reddy, K. M.; Feris, K.; Bell, J.; Wingett, D. G.; Hanley, C.;  
Punnoose, A. Selective toxicity of zinc oxide nanoparticles to  
prokaryotic and eukaryotic systems. *Appl. Phys. Lett.* **2007**, *90*,  
2139021–2139023. 816
- (35) Premanathan, M.; Karthikeyan, K.; Jeyasubramanian, K.;  
Manivannan, G. Selective toxicity of ZnO nanoparticles toward  
Gram-positive bacteria and cancer cells by apoptosis through lipid  
peroxidation. *Nanomedicine* **2011**, *7* (2), 184–192. 820
- (36) Khosla, H.; Seche, W.; Ammerman, D.; Elyahoodayan, S.;  
Caputo, G. A.; Hettlinger, J.; Amini, S.; Feng, G. Development of  
antibacterial neural stimulation electrodes via hierarchical surface  
restructuring and atomic layer deposition. *Sci. Rep.* **2023**, *13* (1),  
No. 19778. 825
- (37) Shepherd, R. K.; Villalobos, J.; Burns, O.; Nayagam, D. A. X.  
The development of neural stimulators: a review of preclinical safety  
and efficacy studies. *J. Neural Eng.* **2018**, *15* (4), No. 041004. 828
- (38) Cogan, S. F. Neural stimulation and recording electrodes. *Annu.*  
*Rev. Biomed. Eng.* **2008**, *10* (1), 275–309. 830
- (39) Bauer, A. W.; Kirby, W. M. M.; Sherris, J. C.; Turck, M.  
Antibiotic Susceptibility Testing by a Standardized Single Disk  
Method. *Am. J. Clin. Pathol.* **1966**, *45* (4), 493–496. 833
- (40) Tay, Y. Y.; Li, S.; Sun, C.; Chen, P. Size dependence of Zn 2p  
3/2 binding energy in nanocrystalline ZnO. *Appl. Phys. Lett.* **2006**, *88*  
(17), No. 2198821, DOI: 10.1063/1.2198821. 836
- (41) Kamarulzaman, N.; Kasim, M. F.; Chayed, N. F. Elucidation of  
the highest valence band and lowest conduction band shifts using XPS  
for ZnO and Zn<sub>0.99</sub>Cu<sub>0.01</sub>O band gap changes. *Results Phys.* **2016**, *6*,  
217–230. 840
- (42) Das, J.; Pradhan, S. K.; Sahu, D. R.; Mishra, D. K.; Sarangi, S.  
N.; Nayak, B. B.; Verma, S.; Roul, B. K. Micro-Raman and XPS  
studies of pure ZnO ceramics. *Phys. B* **2010**, *405* (10), 2492–2497. 843
- (43) Moulder, J. F.; Stickle, W. F.; Sobol, P. E.; Bomben, K. D.  
*Handbook of X-ray Photoelectron Spectroscopy*; Perkin-Elmer Corpo-  
ration, 1992. 846
- (44) Frankcombe, T. J.; Liu, Y. Interpretation of Oxygen 1s X-ray  
Photoelectron Spectroscopy of ZnO. *Chem. Mater.* **2023**, *35* (14),  
5468–5474. 849
- (45) Samsonov, G. V. *The Oxide Handbook*; Springer Science &  
Business Media, 2013. 851
- (46) Roiron, C.; Wang, C.; Zenyuk, I. V.; Atanassov, P. Oxygen 1s  
X-ray Photoelectron Spectra of Iridium Oxides as a Descriptor of the  
Amorphous–Rutile Character of the Surface. *J. Phys. Chem. Lett.* **2024**, *15* (45), 11217–11223. 855
- (47) Illy, B. N.; Cruickshank, A. C.; Schumann, S.; Da Campo, R.;  
Jones, T. S.; Heutz, S.; McLachlan, M. A.; McComb, D. W.; Riley, D.  
J.; Ryan, M. P. Electrodeposition of ZnO layers for photovoltaic  
applications: controlling film thickness and orientation. *J. Mater.*  
*Chem.* **2011**, *21* (34), 12949–12957. 860
- (48) Cruickshank, A. C.; Tay, S. E. R.; Illy, B. N.; Da Campo, R.;  
Schumann, S.; Jones, T. S.; Heutz, S.; McLachlan, M. A.; McComb,  
D. W.; Riley, D. J.; Ryan, M. P. Electrodeposition of ZnO 863

864 Nanostructures on Molecular Thin Films. *Chem. Mater.* **2011**, 23  
865 (17), 3863–3870.

866 (49) Manzano, C. V.; Alegre, D.; Caballero-Calero, O.; Alén, B.;  
867 Martín-González, M. S. Synthesis and luminescence properties of  
868 electrodeposited ZnO films. *J. Appl. Phys.* **2011**, 110 (4), No. 043538.

869 (50) Xing, L.; Vatamanu, J.; Smith, G. D.; Bedrov, D. Nano-  
870 patterning of Electrode Surfaces as a Potential Route to Improve the  
871 Energy Density of Electric Double-Layer Capacitors: Insight from  
872 Molecular Simulations. *J. Phys. Chem. Lett.* **2012**, 3 (9), 1124–1129.

873 (51) Vatamanu, J.; Cao, L.; Borodin, O.; Bedrov, D.; Smith, G. D.  
874 On the Influence of Surface Topography on the Electric Double Layer  
875 Structure and Differential Capacitance of Graphite/Ionic Liquid  
876 Interfaces. *J. Phys. Chem. Lett.* **2011**, 2 (17), 2267–2272.

877 (52) Vatamanu, J.; Hu, Z.; Bedrov, D.; Perez, C.; Gogotsi, Y.  
878 Increasing Energy Storage in Electrochemical Capacitors with Ionic  
879 Liquid Electrolytes and Nanostructured Carbon Electrodes. *J. Phys.*  
880 *Chem. Lett.* **2013**, 4 (17), 2829–2837.